

Canadian COVID-19 Wastewater Coalition

Webinar series – Tuesday, December 1, 2020



Inter-Laboratory Study Outcomes & Implications

11:30 a.m. to 12:55 p.m. EST



CWN Webinars

Connecting water professionals to decision-ready knowledge



COVID-19 WASTEWATER COALITION

A national collaboration of municipal utilities, researchers, public health agencies and government with a shared goal of protecting public health from COVID-19

[COALITION UPDATES](#)[NEWS STORIES](#)[INTER-LAB STUDY](#)[REGIONAL HUBS](#)[RESOURCES](#)[GET INVOLVED](#)

“Helping Canada to better address and assess what is needed and what the reliable use of the technique could look like is the fundamental rationale for the Canadian COVID-19 Wastewater Coalition.”

Steve E. Hrudey et al., June 2020

Royal Society of Canada COVID-19 Series | Publication #23

COVID-19 Wastewater Coalition Research

Draft Framework April 23, 2020

Main end users of research outcomes

- Public health surveillance
- Emergency Operations Centres
- Health regulators

Decision Application 1

SARS-CoV-2 wastewater surveillance as an aggregated indicator of community COVID-19, including spread & asymptomatic presence

Decision Application 2

SARS-CoV-2 infectivity and exposure via wastewater, including risk assessment, environmental fate & transport

Main end users of research outcomes

- Public health
- Utilities and municipalities
- Environmental and systems regulators
- Occupational health groups

Wastewater sampling, processing and preservation/archiving

Analytical techniques –
RT-qPCR, other (Level 2 labs)

Analytical techniques –
genetic, culture methods, other (Level 3 labs)

Wastewater system characterization and modelling to enable appropriate interpretation and connection of results to community.

Epidemiological and surveillance modelling to apply results to inform trends and community prevalence analysis

Wastewater system characterization to link results to risk assessments and transport assessments

Application of results to appropriate risk assessment considerations

Research Elements

Effective and transparent communication and positioning of results to better support evidence-informed decisions and public engagement

How can SARS-CoV-2 sewage surveillance best support public health decisions?

- Reflecting asymptomatic and pre-symptomatic in addition to symptomatic individuals?
- Providing an efficient pooled sample?
- Tracking community trends?
- Potential to detect low levels of infection from communities or facilities (sentinel)?
- Potential to better understand spread within a community (support epidemiology)?

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Advancing confidence in analytical results

- A rapidly evolving area globally
- Data reliability appropriate to public health decisions
- Immediate needs (pressure for fast turnaround)
- Advance collectively toward consensus approaches
- Improve and accelerate collective understanding, capacity and expertise in Canada

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Inter-Laboratory Study – Participating Laboratories

- Public Health Agency of Canada – National Microbiology Laboratory
- Public Health Laboratory – Alberta Precision Laboratory;
University of Alberta
- BC Centre for Disease Control; University of British Columbia
- Toxicology Centre; University of Saskatchewan
- University of Ottawa
- University of Waterloo
- Great Lakes Institute for Environmental Research;
University of Windsor
- Polytechnique Montréal



Canadian
Water
Network

COVID-19 WASTEWATER COALITION

Inter-laboratory Study Outcomes & Implications

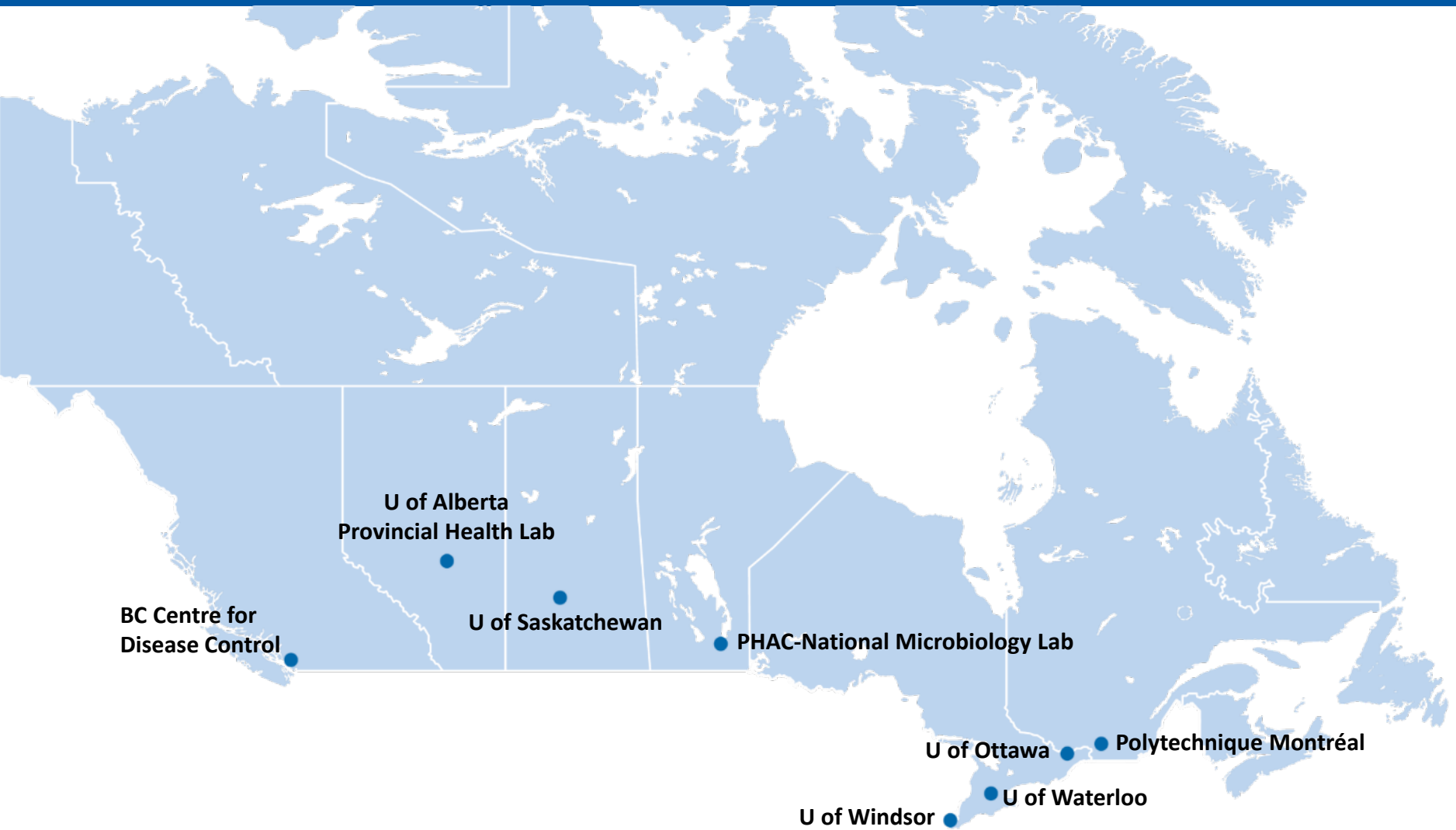
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Inter-Laboratory Study

- ❁ Laboratory capabilities were solicited by the CWN Coalition
- ❁ Discussions held with labs indicating a method was operational
- ❁ Ultimately 7 laboratories & National Microbiology Lab involved
- ❁ Study design was developed by a sub-group from these labs
- ❁ A 12-litre wastewater grab sample was provided by Winnipeg at a time when cases were low (~85 active cases in population of 750,000)
- ❁ NML shipped each lab a set of wastewater samples with no spike, low spike and high spike

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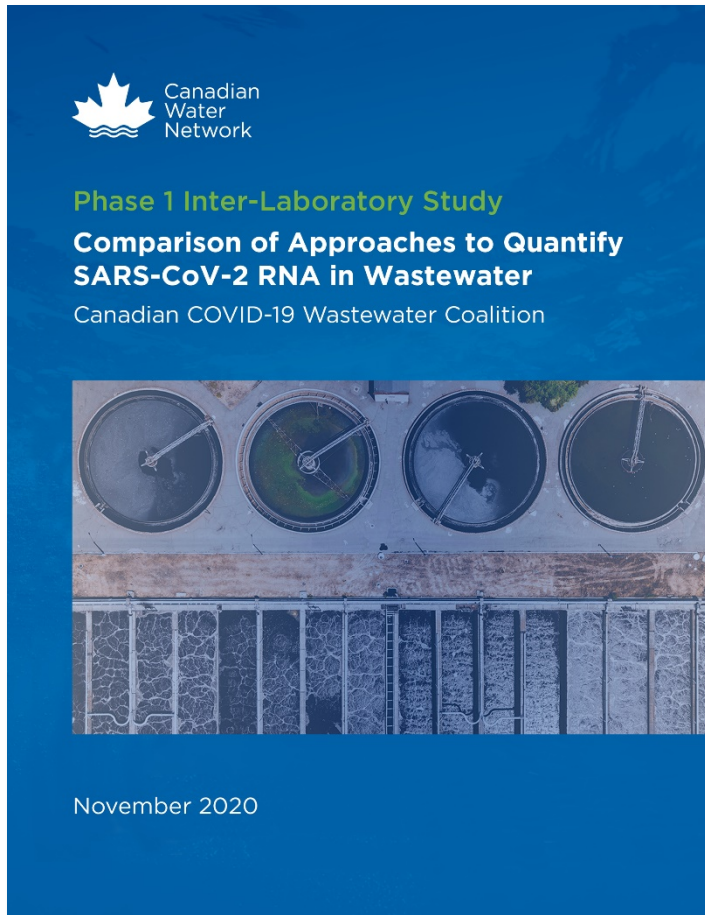
Inter-Laboratory Study – Participating Laboratories



Phase 1 Inter-Laboratory Study

- ❁ Characterize the inter- and intra- laboratory variability associated with results from the testing of SARS-CoV-2 using RT-qPCR after extraction from a common wastewater matrix
- ❁ Inter-laboratory variability in results could be due to differences in the wastewater sample pre-treatment method
- ❁ Recognize the diversity of protocols and supply chain limitations
- ❁ Leverage existing expertise and capacity

Available at: cwn-rce.ca/covid-19-wastewater-coalition



Who should read the report?

- ❁ Public health leaders seeking to understand the potential (and limitations) of wastewater surveillance
- ❁ Decision makers considering the feasibility of wastewater surveillance programs
- ❁ Laboratories in the process of developing or adapting SARS-CoV-2 RT-qPCR methods to various wastewater matrices



Public Health
Agency of Canada

Agence de la santé
publique du Canada

Canada

Preparation of wastewater reference material for CWN/PHAC interlab study

Dr. Chand S. Mangat

Dr. Michael R. Mulvey

Dr. James Brooks

2020-12-2

PROTECTING AND EMPOWERING CANADIANS
TO IMPROVE THEIR HEALTH



PHAC-NML COVID-19 Activities

- Clinical laboratory diagnosis and surveillance
 - “In-country” procurement/development coordination for reagents and supplies for COVID testing and research in Canada
 - Provincial / Territorial surge capacity
 - NRI mobile testing deployment
 - POC test validation and deployment
 - Evaluation of decontamination procedures for disinfection of surfaces and equipment
 - Seroprevalence survey
 - Whole viral sequencing of clinical samples (CanCoGen)
 - Development and investigation of vaccines
- Modelling strategies to predict SARS-CoV2 transmissions and clusters
- Research and Development
 - Development of in vitro and animal models for growth and study of SARS-CoV-2
 - Recombinant SARS-CoV-2 and seasonal coronavirus proteins for downstream analysis and reagent use
- **Wastewater surveillance**

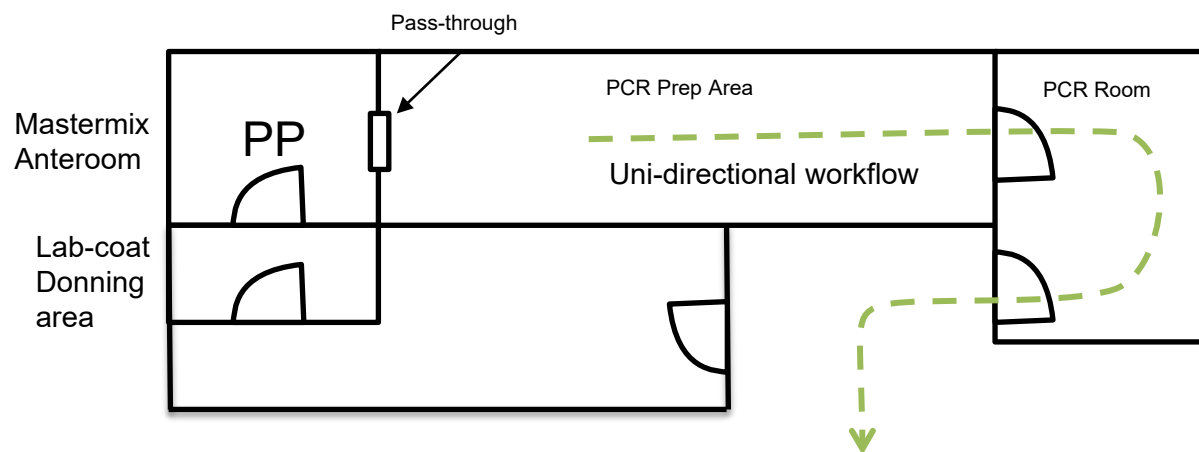
CWN Interlab Study – Sample collection and distribution

- 8 participating labs
- Spiking material was prepared prior to arrival of grab sample in a separate area
 - gamma inactivated SARS-CoV-2 (NML)
 - Quantified by RT-ddPCR (Bio-rad N1/N2 duplex)
 - Hcov 229E (Lilly Pang, AB Precision lab)
- 12 L grab sample from Winnipeg North End treatment plant in HDPE bottles (August 31st)
 - Bottles were validated to not deplete samples of PMMV or SARS-CoV-2 over 12 days.
 - Shipped on ice, arrived at lab within 20 minutes of collection
- 3 x 100 mL samples prepared for each lab
 - Unspiked (N)
 - Spiked low (A, 18 copies/ mL)
 - Spiked high (B, 1800 copies/mL)

CWN Interlab Study – Sample collection and distribution

- Un-spiked samples were processed first in a separate BSC, all unspiked bottles were closed and stored prior to spiking
- Bottles randomized prior to packing
- Sample packaging was validated to maintain temperature for ($\sim 2^{\circ}\text{C}$) for two days when stored at ambient temperature.
- Sample shipped UN3373 Cat. B
- Samples collected, spiked and shipped the same day to mimic a sampling event.

NML PCR diagnostics area used for WW



Wastewater partners

- PHAC
 - Dr. Chand Mangat
 - Dr. Michael R. Mulvey
 - Dr. James Brooks
 - Dr. David Champredon
 - Aamir Fazil
 - Jayson Shurgold
 - Howard Swerdfeger
 - Dr. Aboubakar Mounchili
 - Dr. Guillaume Poliquin
 - Dr. Paul Sandstrom
 - Dr. Adrienne Myers
 - **Dr. Michael Becker**
 - Dr. Heidi Wood
 - Dr. Michael Drebot
 - **Dr. Nathalie Bastien**
 - Dr. Patrick Chong
 - Dr. Garrett Westmacott
 - Dr. Chrystal Landgraff
 - Dr. Celine Nadone

Thank you for your time and consideration.

- **Jade Daigle**
 - **Ravinder Lidder**
 - **Dave Spreitzer**
 - **Stacie Langner**
- PHAC-OCSO/OSSI
 - Dr. Pascal Michel
 - Mette Cornelisse
- STATCAN
 - Audra Nagasawa
 - Thac Dung (TD) Nguyen
- ECCC
 - Shirley Anne Smythe
 - Steven Teslic
- **Canadian Water Network**
 - **Bernadette Conant**
 - **Dr. Steven Hrudehy**
 - **Dr. Alex Chik**
 - **CWN members**

Phase 1 Inter-Laboratory Study

Comparison of Approaches to Quantify SARS-CoV-2 RNA in Wastewater

Canadian COVID-19 Wastewater Coalition



November 2020

Available at cwn-rce.ca

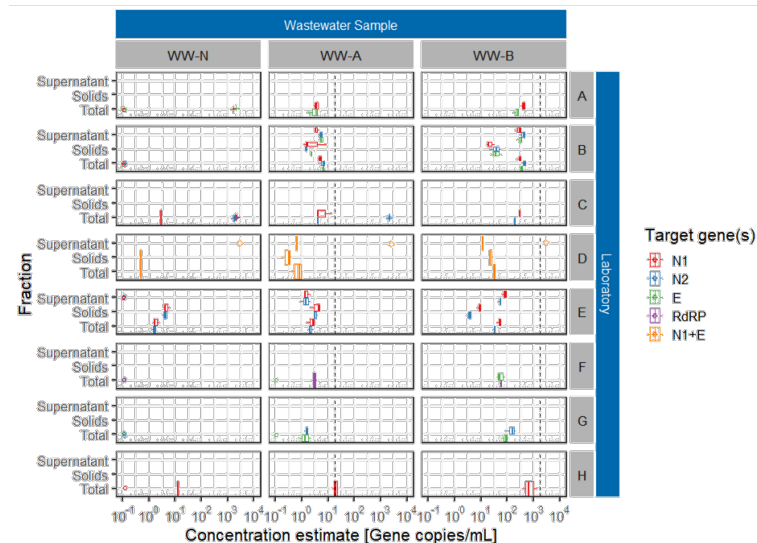


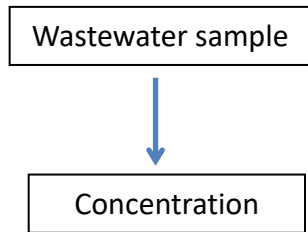
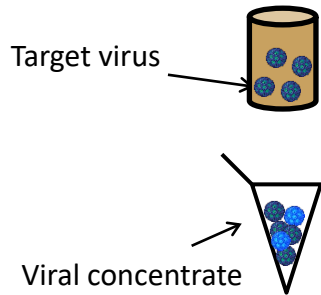
Table 3 SARS-CoV-2 \log_{10} concentration estimates observed in spiked Winnipeg wastewater samples. Standard deviation (SD) and coefficient of variation (COV; Canchola et al., 2017) statistics were only calculated where all three aliquots of each condition yielded quantifiable SARS-CoV-2 concentration estimates.

Laboratory	Target gene(s)	WW-A			WW-B		
		Mean	SD	COV (%)	Mean	SD	COV (%)
A	E	0.47	0.26	64.9	2.37	0.15	35.4
	N1	0.55	0.15	36.2	2.60	0.14	32.8
B	E	0.81	0.08	18.0	2.54	0.09	19.8
	N1	0.69	0.11	24.8	2.47	0.13	30.0
C	N2	0.77	0.19	47.0	2.63	0.13	30.0
	N1	0.77	0.48	152.7	2.48	0.01	3.3
D	N2	-	-	-	2.28	0.04	9.1
	N1+E	-0.18	0.32	85.4	1.50	0.04	9.6
E	N1	0.37	0.20	49.7	1.70	0.13	31.5
	N2	0.33	0.18	42.9	1.51	0.06	14.8
F	E	-	-	-	1.75	0.05	12.1
	RdRP	-	-	-	1.75	0.05	12.1
G	N2	-	-	-	2.14	0.26	64.2
	N1	1.28	0.17	41.8	2.84	0.40	114.3

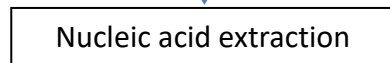
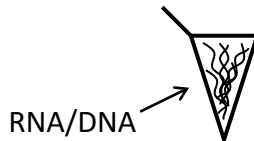
Chik et al., for submission to
Journal of Environmental Sciences

Overview of Methods

Sample Preparation



Nucleic Acid Extraction



Detection and Quantification

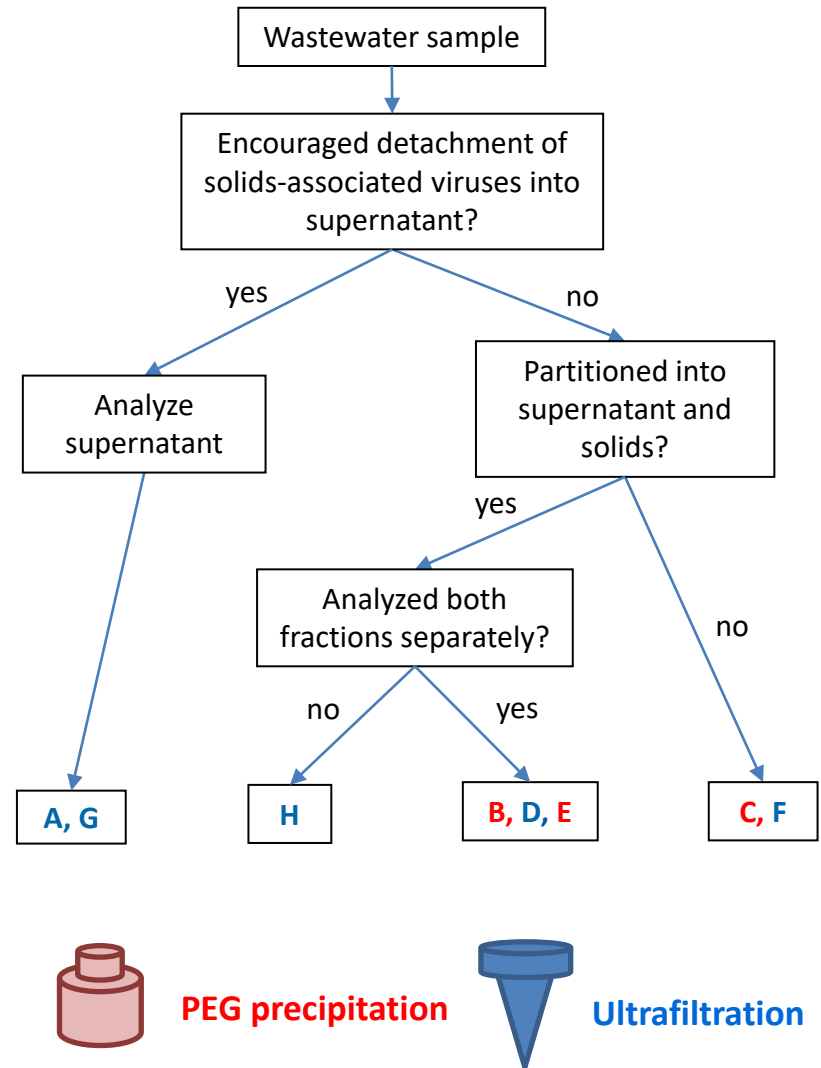
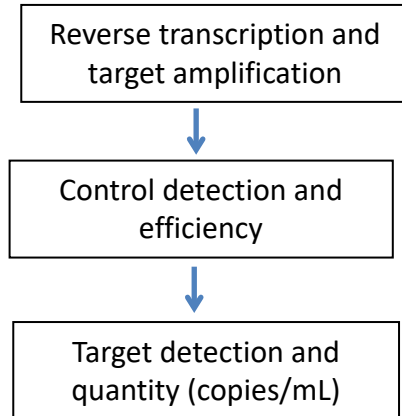
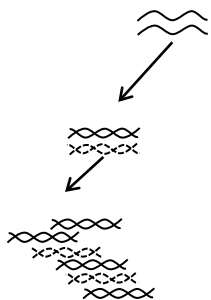
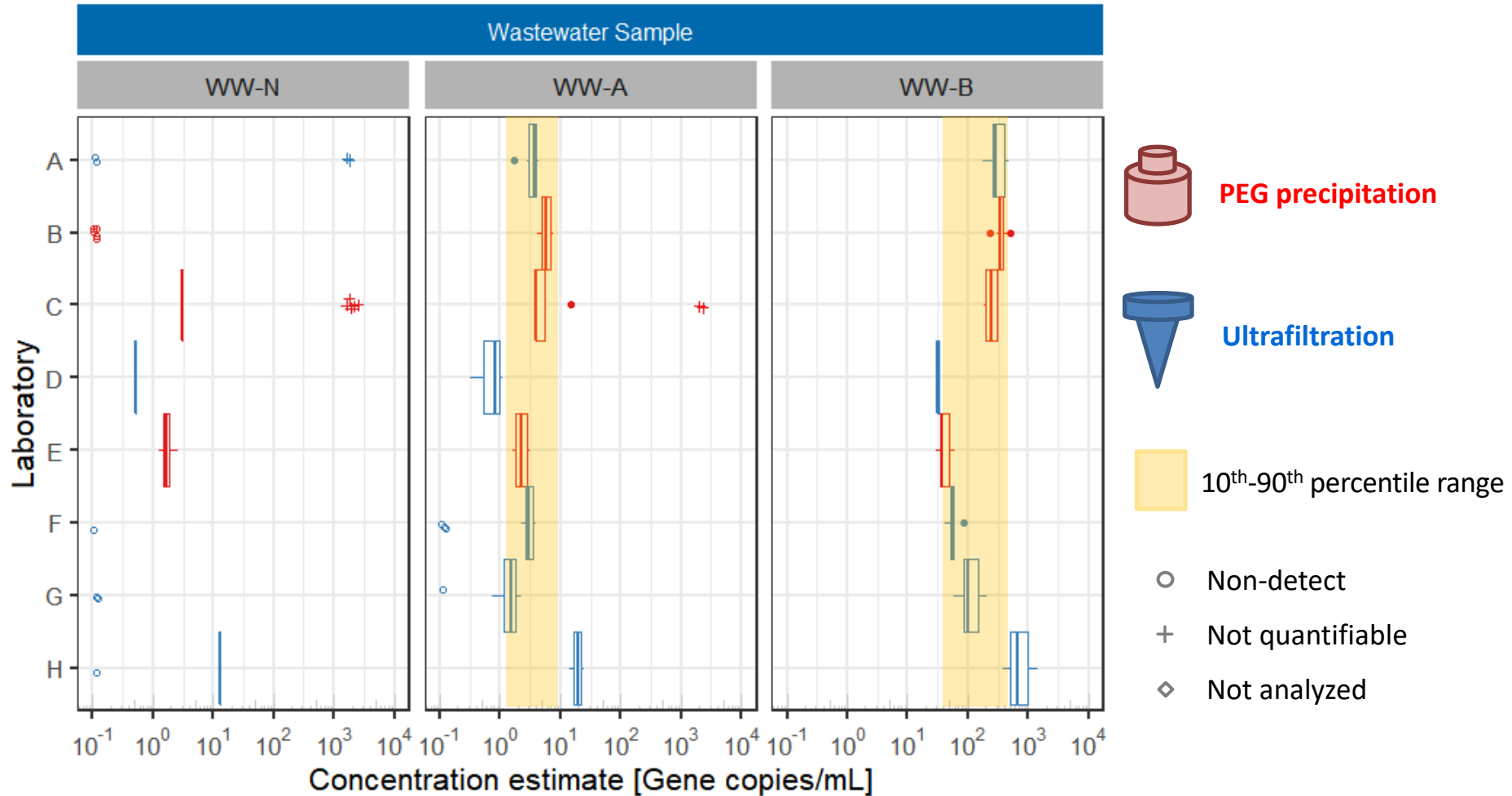
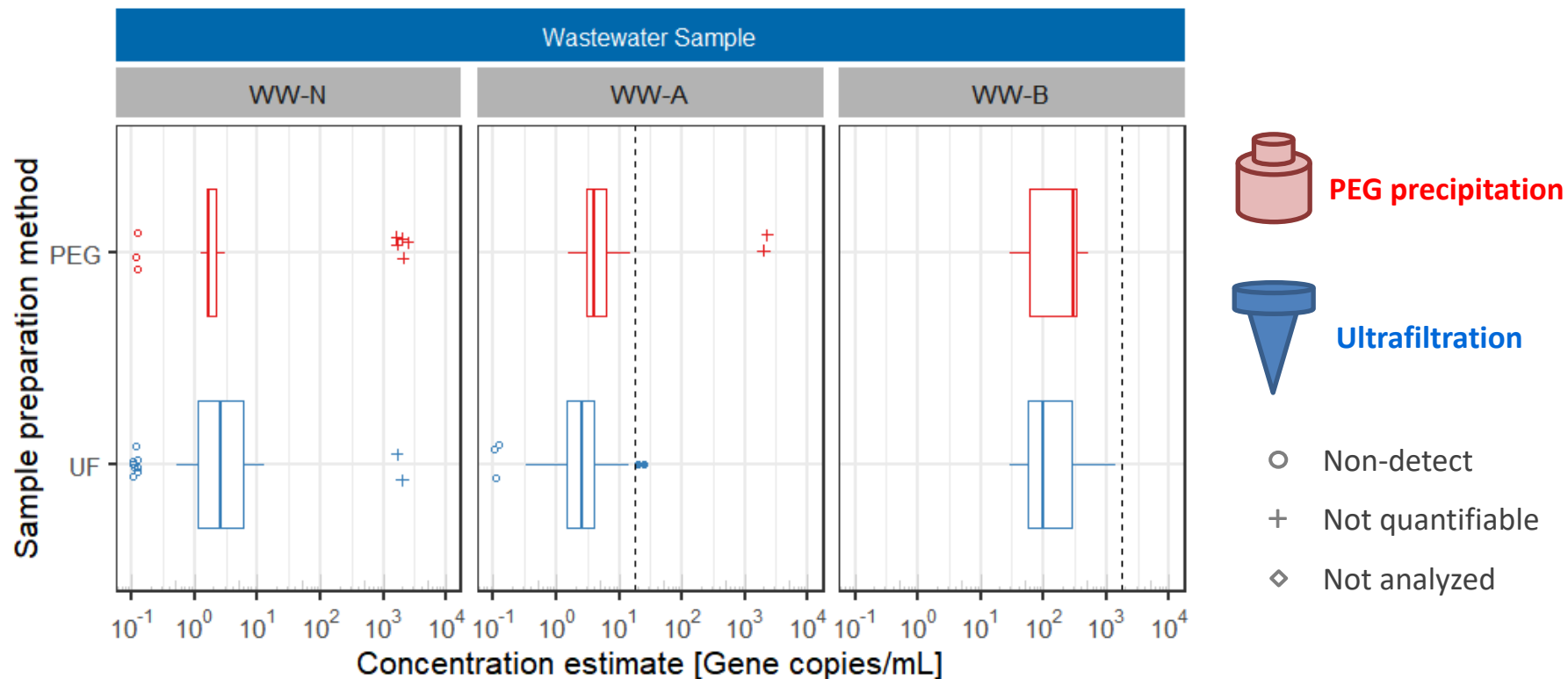


Figure adapted from Iker et. al. 2016 with modifications by M. Glier

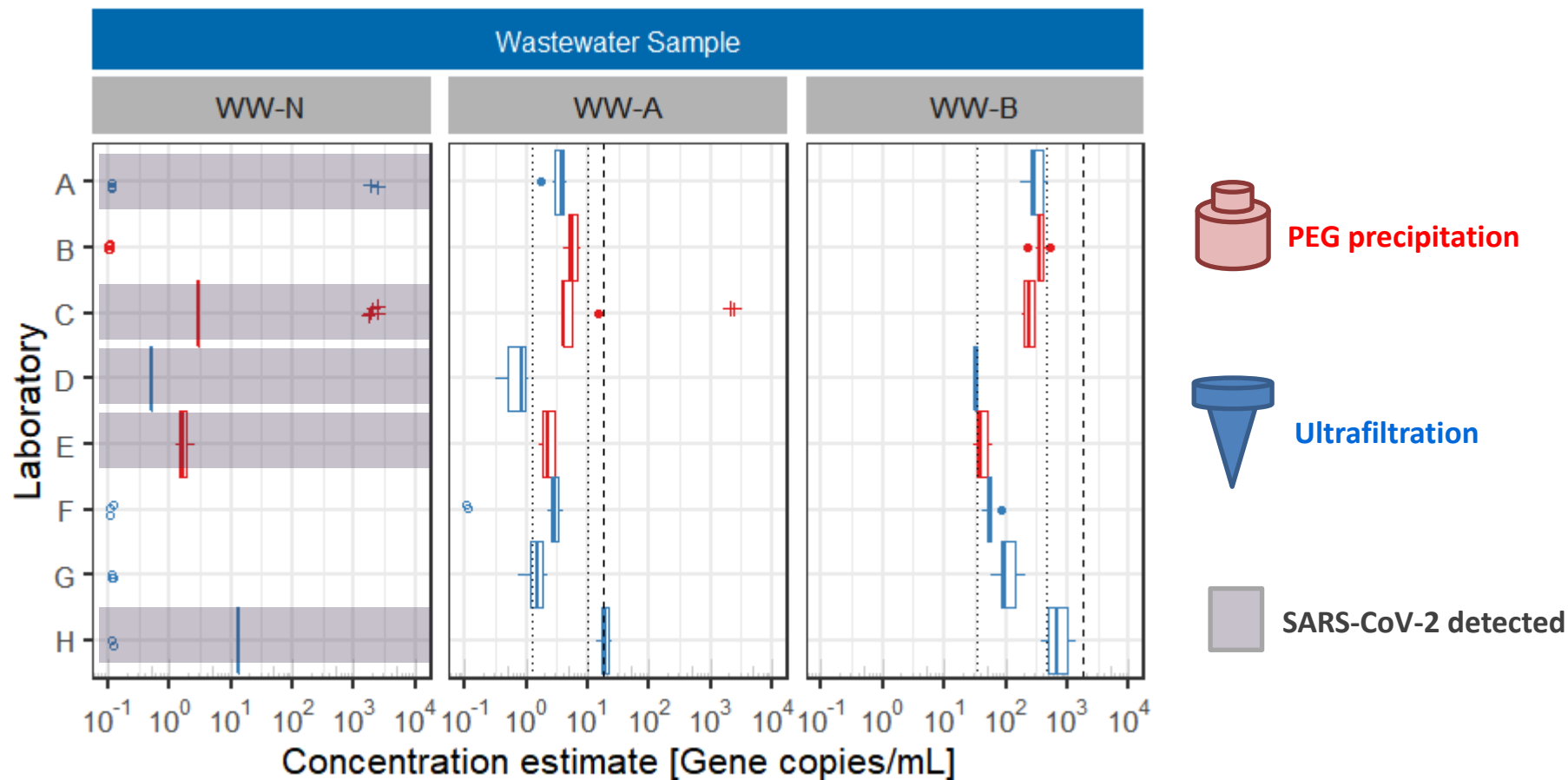
SARS-CoV-2 RNA measured in Winnipeg wastewater matrix



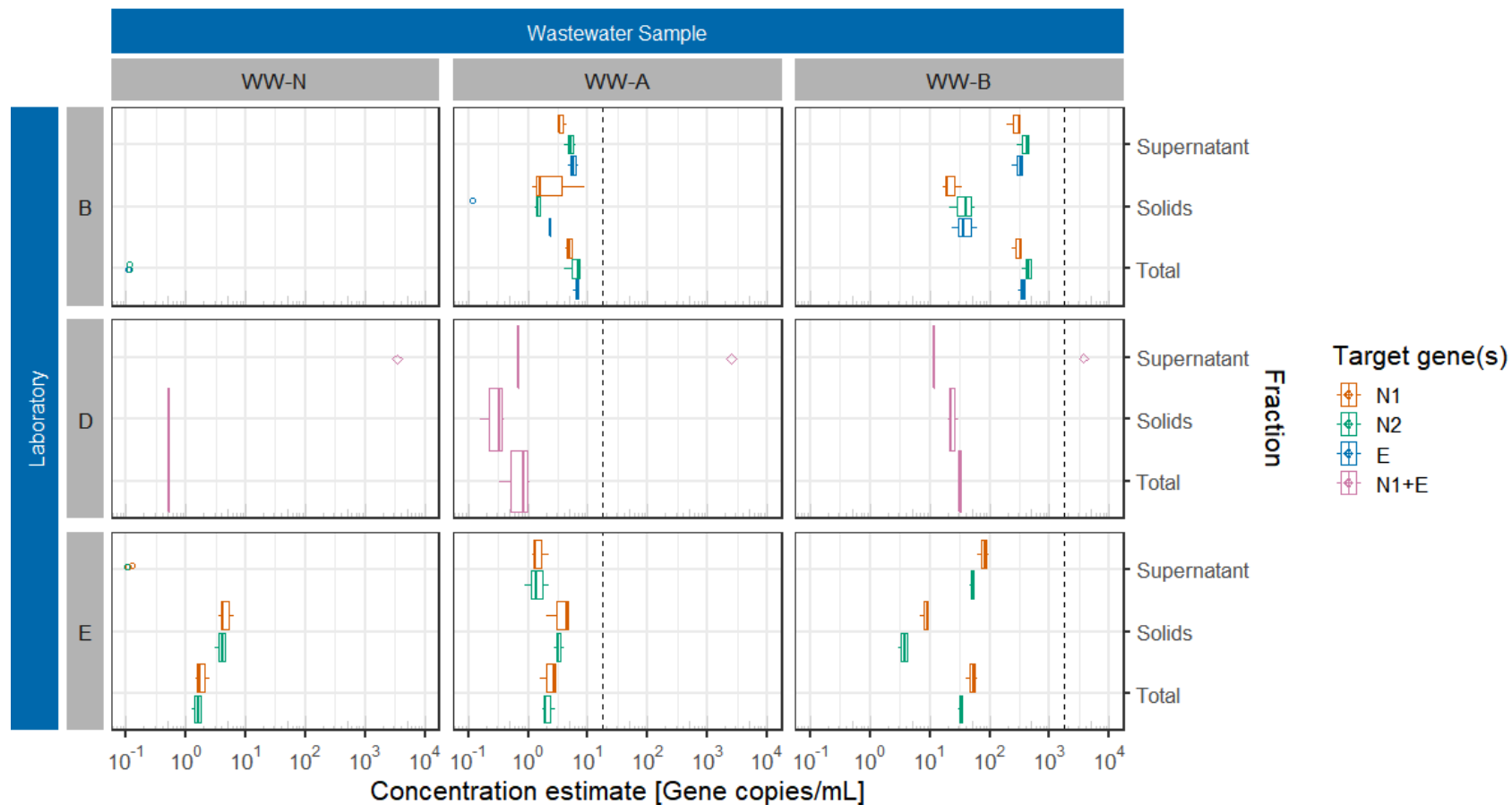
Comparison across sample preparation methods



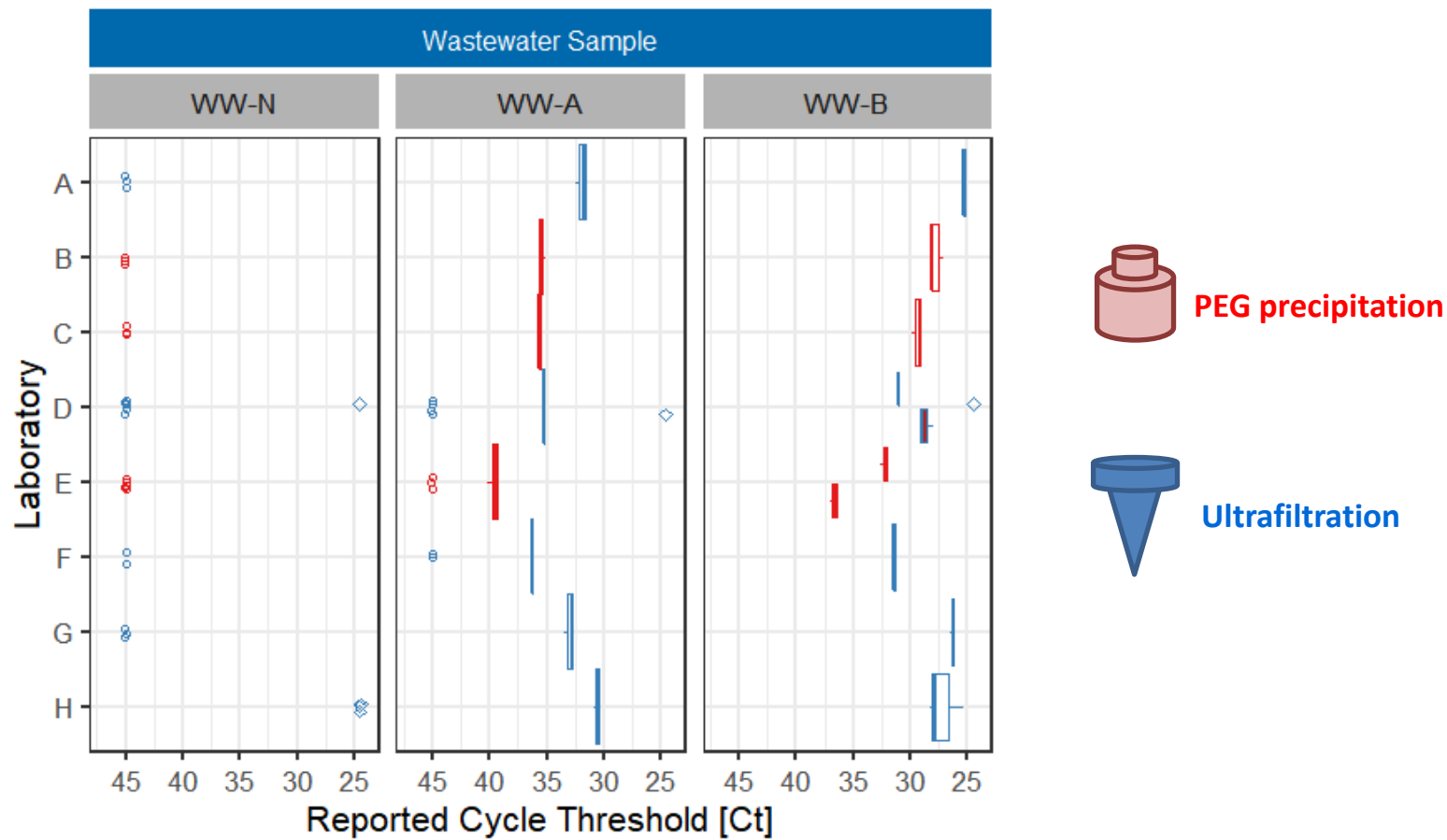
SARS-CoV-2 RNA measured in Winnipeg wastewater matrix



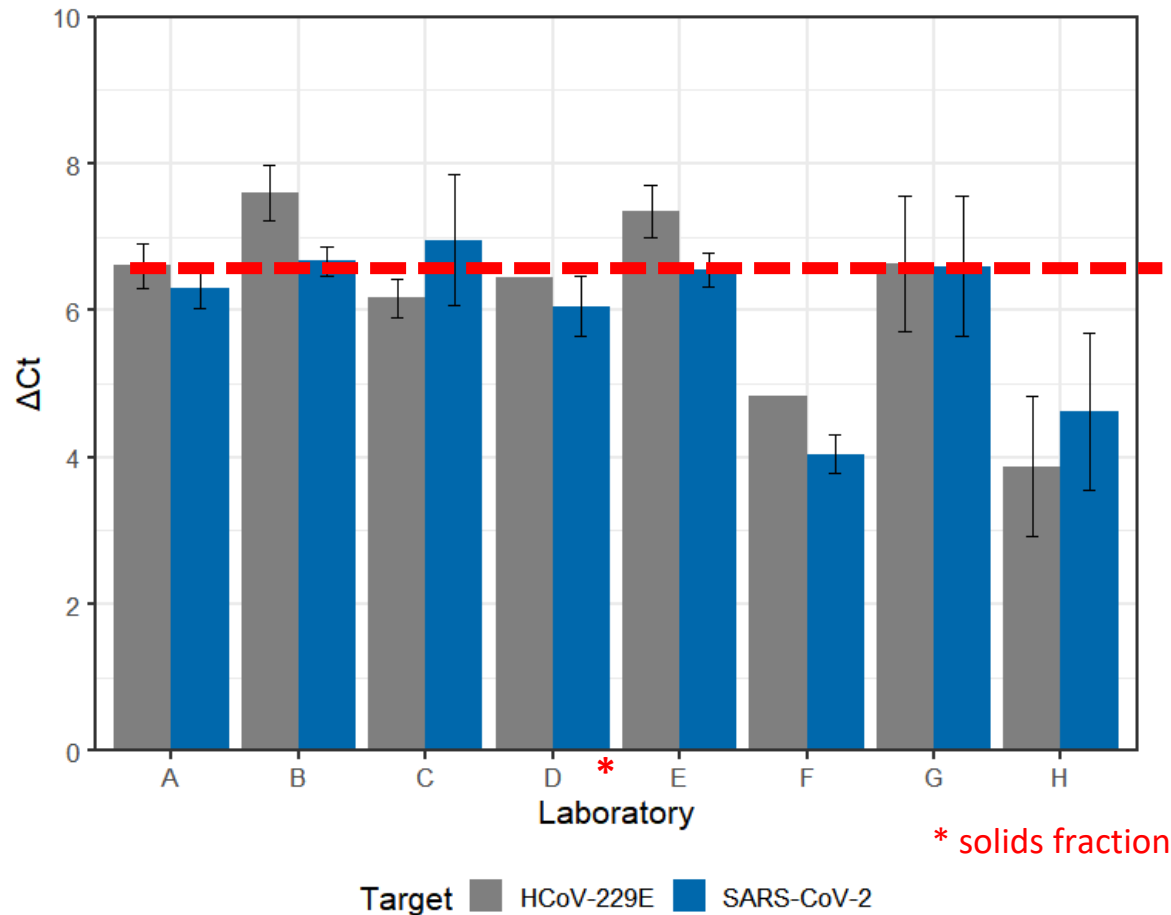
SARS-CoV-2 partitioning



HCoV-229E RNA measured in Winnipeg wastewater matrix



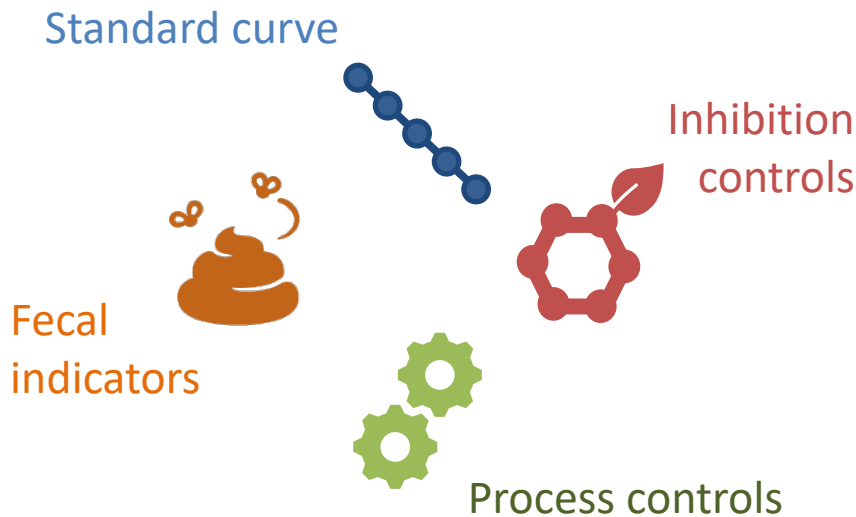
Difference between high and low spike cycle thresholds for both spike surrogates



Key findings of the Inter-Laboratory Study

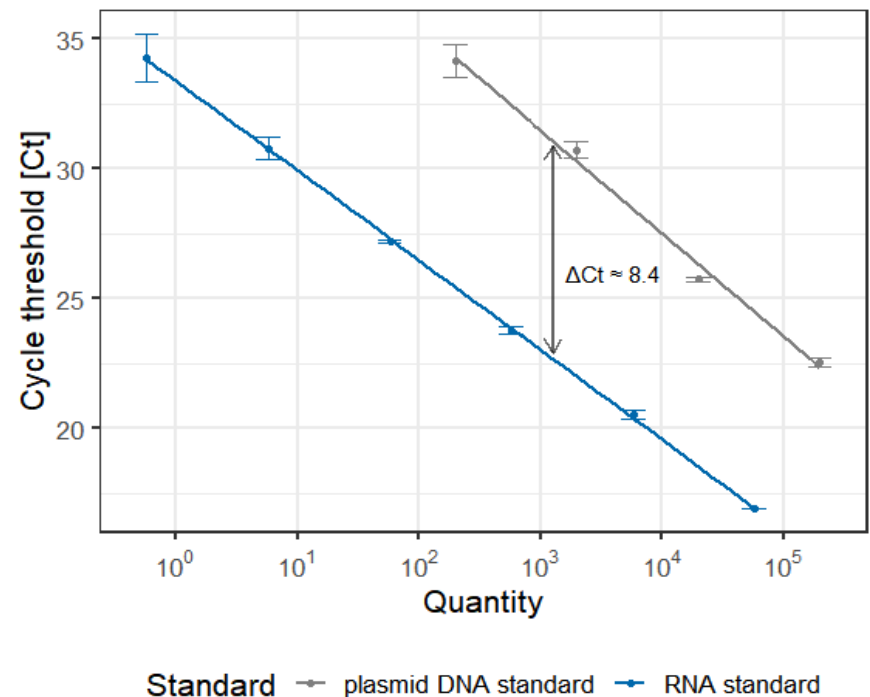
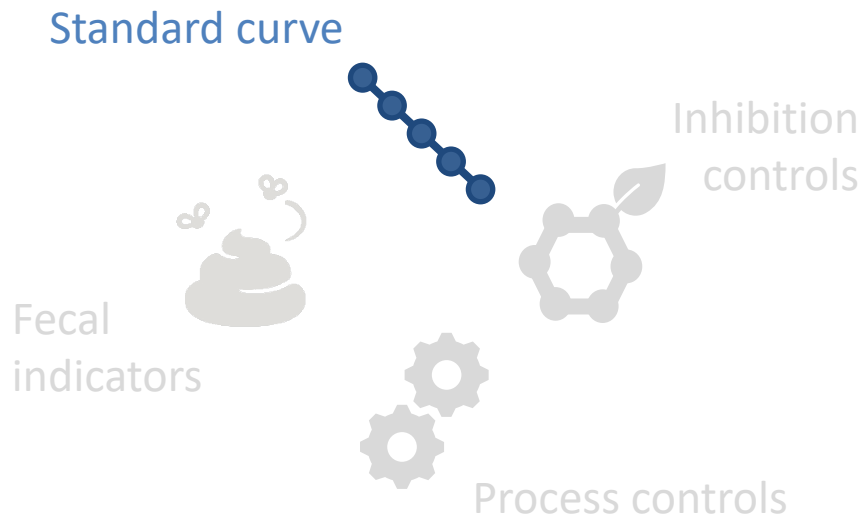
- Highly reproducible results *within* each laboratory
- Comparable results *between* laboratories
 - ✓ High- from low-spike conditions reliably distinguished
- Clear evidence of SARS-CoV-2 surrogates partitioning
- Indication that *in situ* SARS-CoV-2 may be solids-associated
- Correlation between spiked surrogates recovered by each laboratory
 - ✓ Caveats associated with study design highlighted

Challenges to inter-laboratory results comparisons

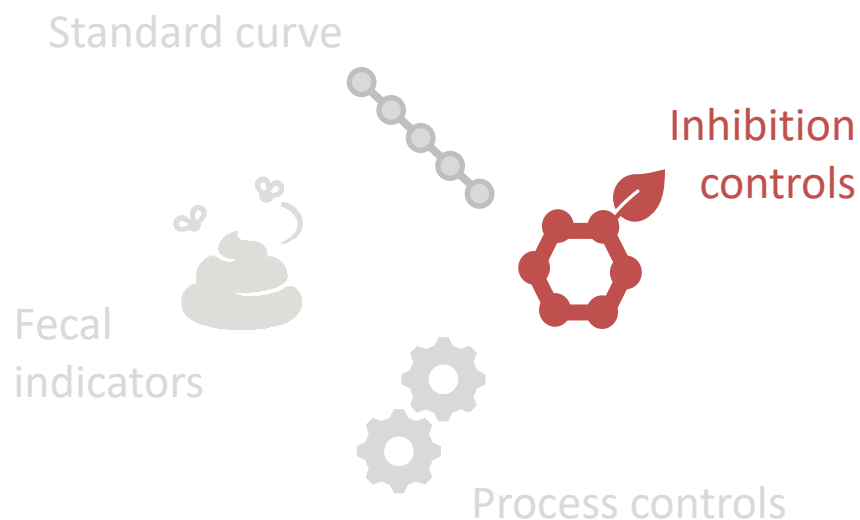


- Coordination between laboratories necessary to ensure apple-to-apple comparisons
- Consensus between laboratories that QA/QC is essential for such data sets

Example of calibration curves based on circular plasmid DNA vs. (linear) RNA standards

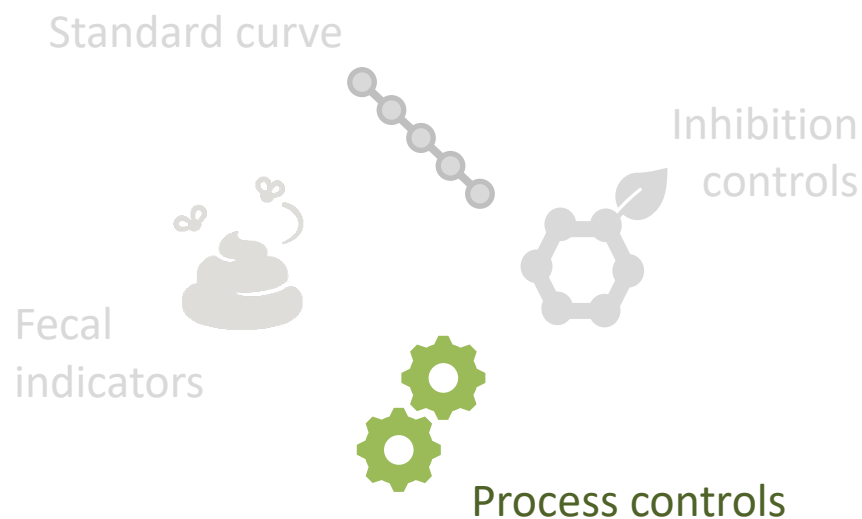


Performing inhibition controls and addressing inhibition if observed



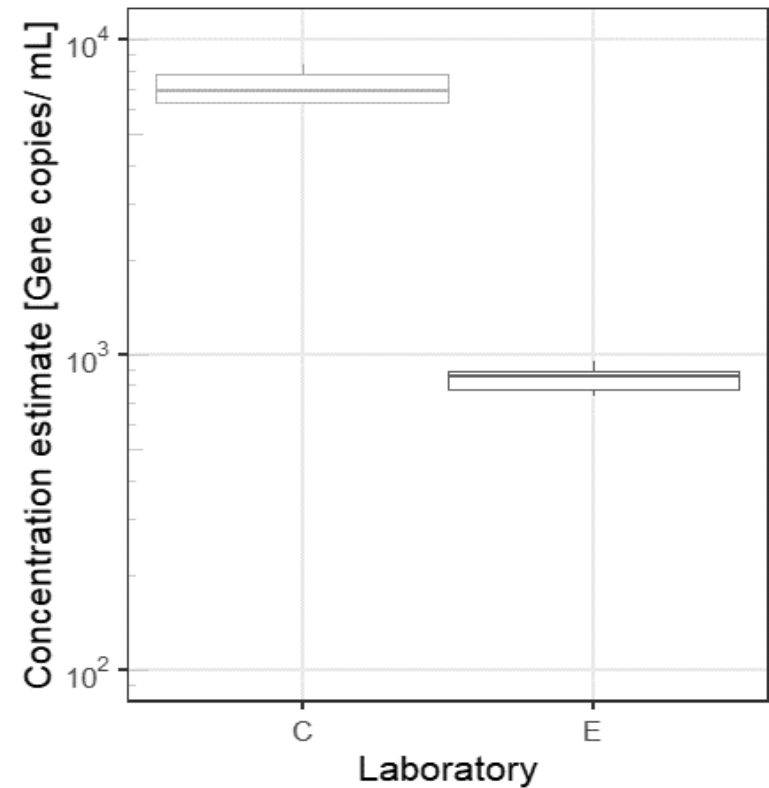
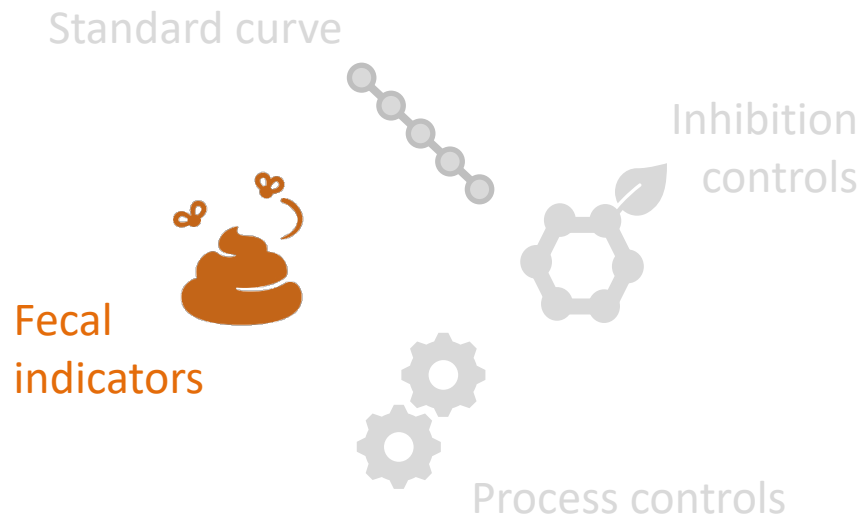
Lab	Indicator	Spike-in at which step	Inhibition when Ct delayed
A	PMMoV		>2
B	Internal positive control	Quantitation	>2
C	MS2 RNA	Quantitation	>2
D			
E	PMMoV		>3
F	West Nile Virus Armoured RNA	RNA extraction	>3-5
G	salmon DNA	RNA extraction	>3
H			

Choice of matrix spikes surrogates and how to perform to estimate recovery



Lab	Surrogate(s) used
A	PPMOV / RNP
B	Armored RNA Quant® human RNase P gene standard
C	Heat-inactivated SARS-CoV-2
D	PMMoV
E	Vesicular stomatitis virus (VSV) inactivated spike
F	Human influenza A (H1N1, viral stock) and human norovirus GII.4 (positive stool sample)
G	Human coronavirus strain 229E (ATCC) with
H	known genetic copies

Fecal virus indicators for supporting detection of wastewater SARS-CoV-2 temporal trends



Building on the Phase 1 Inter-Laboratory Study

- ❁ Bad data is worse than no data
- ❁ Confidence in data is paramount for public health decision making
- ❁ Inter-laboratory study results provide confidence in sample processing and analytical results
- ❁ Phase 2 inter-laboratory study will focus on recovering *in situ* SARS-CoV-2 from a variety of wastewater matrices
- ❁ Value in collective learning

Questions for our panelists

- ❁ What did you get out of participating in the Phase 1 Inter-Laboratory Study?
- ❁ Given your experiences over the last 8 months, including the inter-laboratory study, what are your current thoughts on what is most needed to advance or understand better in the immediate future?

Wastewater monitoring for SARS-CoV-2 in Montréal (QC)

Source :Réseau Canadien de l'Eau

Eyerusalem Goitom¹, Jean-Baptiste Burnet¹, Fernando S. Quete², Sara Matthews^{1,2},
Dominic Frigon², Sarah Dorner¹,

Monitoring SARS-CoV-2 in wastewater



- Québec consortium
 - Monitoring in wastewater (Montréal, Québec, Trois-Rivières,...)
- To identify and understand temporal trends

Opportunities

Tool to complement clinical testing
Early warning?
Facilitate public health decisions?

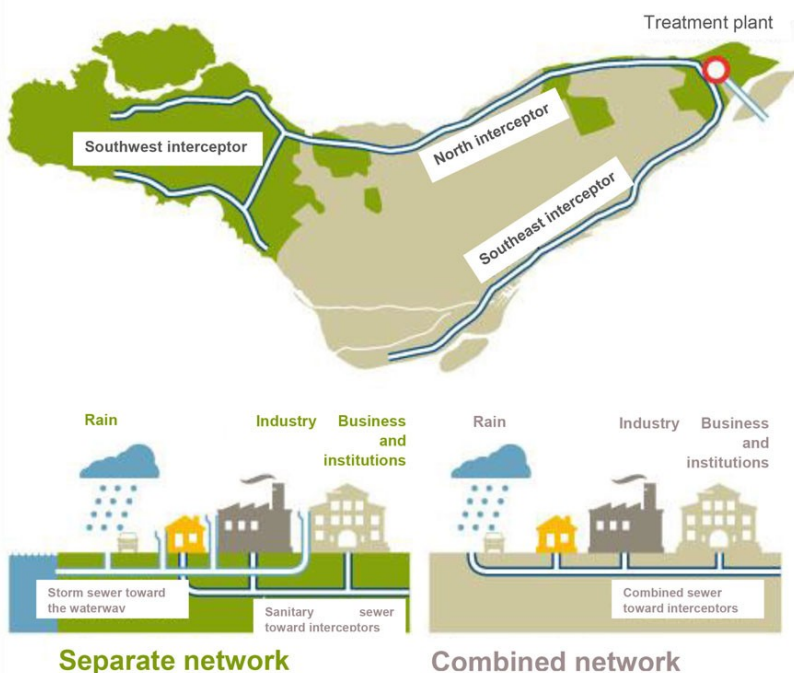
Challenges

Wastewater = complex matrices
What does the virus signal mean?
Sensitivity of the tool?



Montreal Sewer system

The sewer system



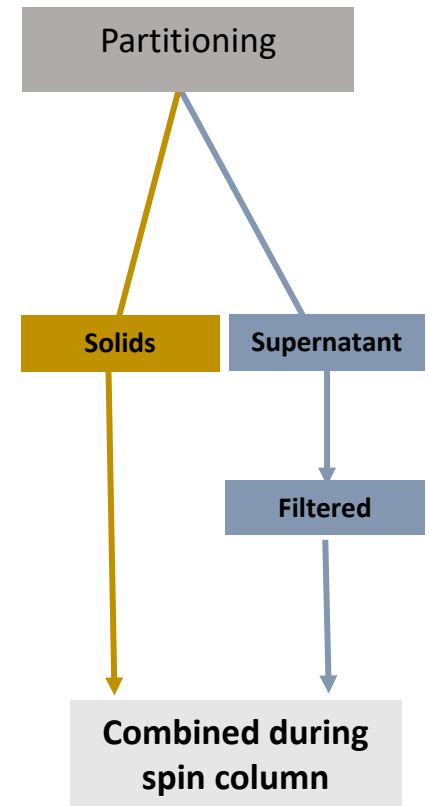
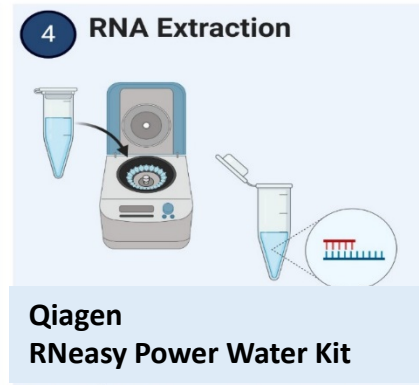
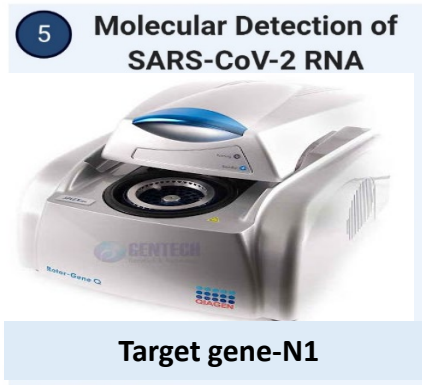
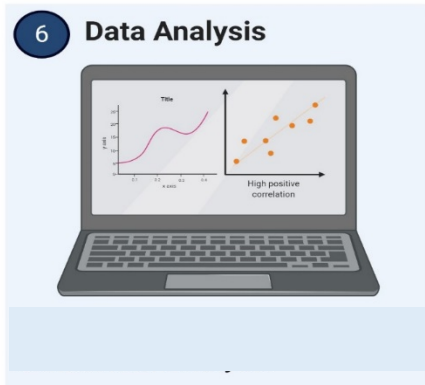
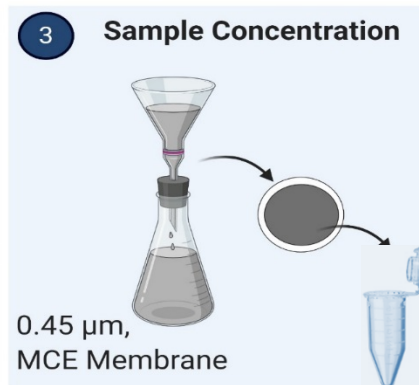
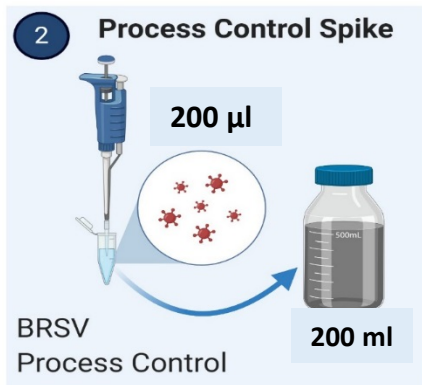
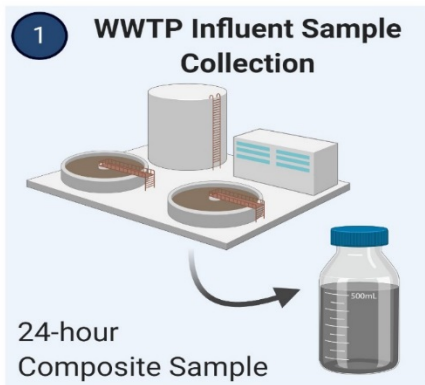
<https://www.canada.ca/en/environment-climate-change/services/water-overview/publications/wastewater-effluent-2015-examination-report.html>

Physiochemical properties of samples in wastewater

Montreal-North-Influent

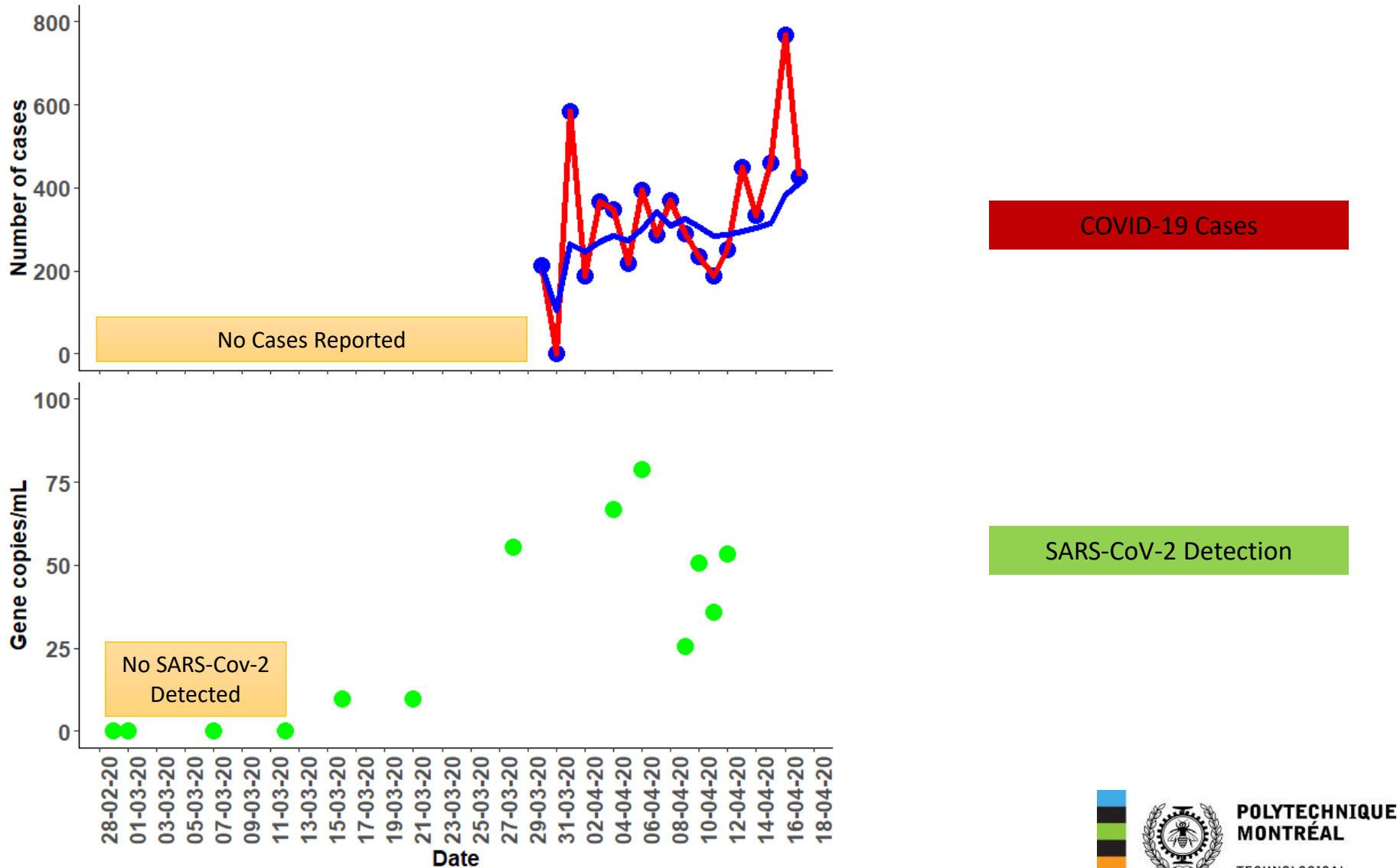
Sampling date	pH	EC (µs/cm)	Turb (NTU)	TDS (ppm)	TSS (mg/L)
2020-02-29	6	NA	66.5	729	148
2020-03-01	6	130.5	66.7	744	NA
2020-03-07	6	126.6	27.10	705	NA
2020-03-12	6	135.5	32.30	808	70
2020-03-16	6	125.3	34.40	709	NA
2020-03-21	6	112.3	43.00	612	NA
2020-03-28	6	NA	29.80	692	52
2020-04-04	6	NA	34.40	550	46
2020-04-06	6	108.8	37.20	571	NA
2020-04-09	6	923.0	64.10	443	NA
2020-04-10	6	881.7	33.60	436	66
2020-04-11	6	981.4	38.90	482	NA
2020-04-12	6	955.1	31.80	479	92
Snowmelt period					

Detection of SARS-CoV-2 RNA in Montreal's wastewater

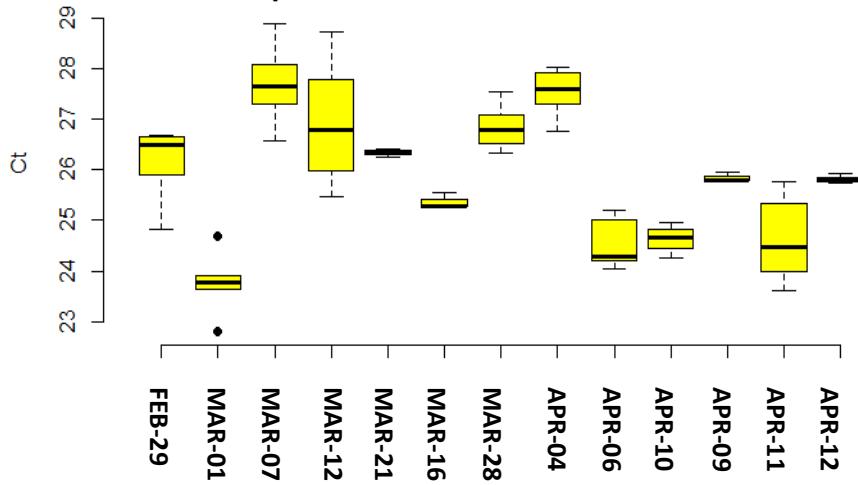


GERM Lab SARS-CoV-2 WBE

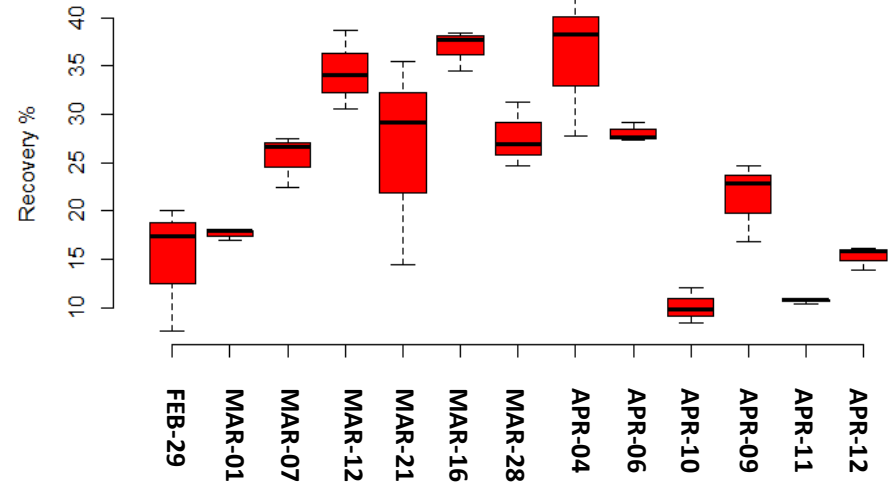
SARS-CoV-2 Detection in Montreal's northern interceptor & Reported Covid-19 cases



Pepper Mottle Virus (PMMoV)



Bovine Respiratory Syncytial Virus (BRSV) Recovery



Conclusions

- ❖ Parallel collaborations increased speed of method development
- ❖ Trends of increases in wastewater were observed prior to increases in reported cases
- ❖ Samples from our archive could be processed after freezing
- ❖ Fecal indicator (PMMoV) are available to normalize SARS-CoV-2 results
- ❖ Recovery indicators (process control) are essential to monitor method performance

Acknowledgements

Canadian Water Network

Bernadette Conant

Alex Chik

Chand Mangat

Liana Kreamer

Lilly Pang

Judy Qiu

UQTR

Elizabeth Grater

François Guillemette

UdeM

Jesse Shapiro

Julie Marleau

Montreal Wastewater Treatment Plant Team

Alexandre Potvin

Carole Fleury

Jean-Claude Deslandes

Jonathan Gagnon

Polytechnique Montreal

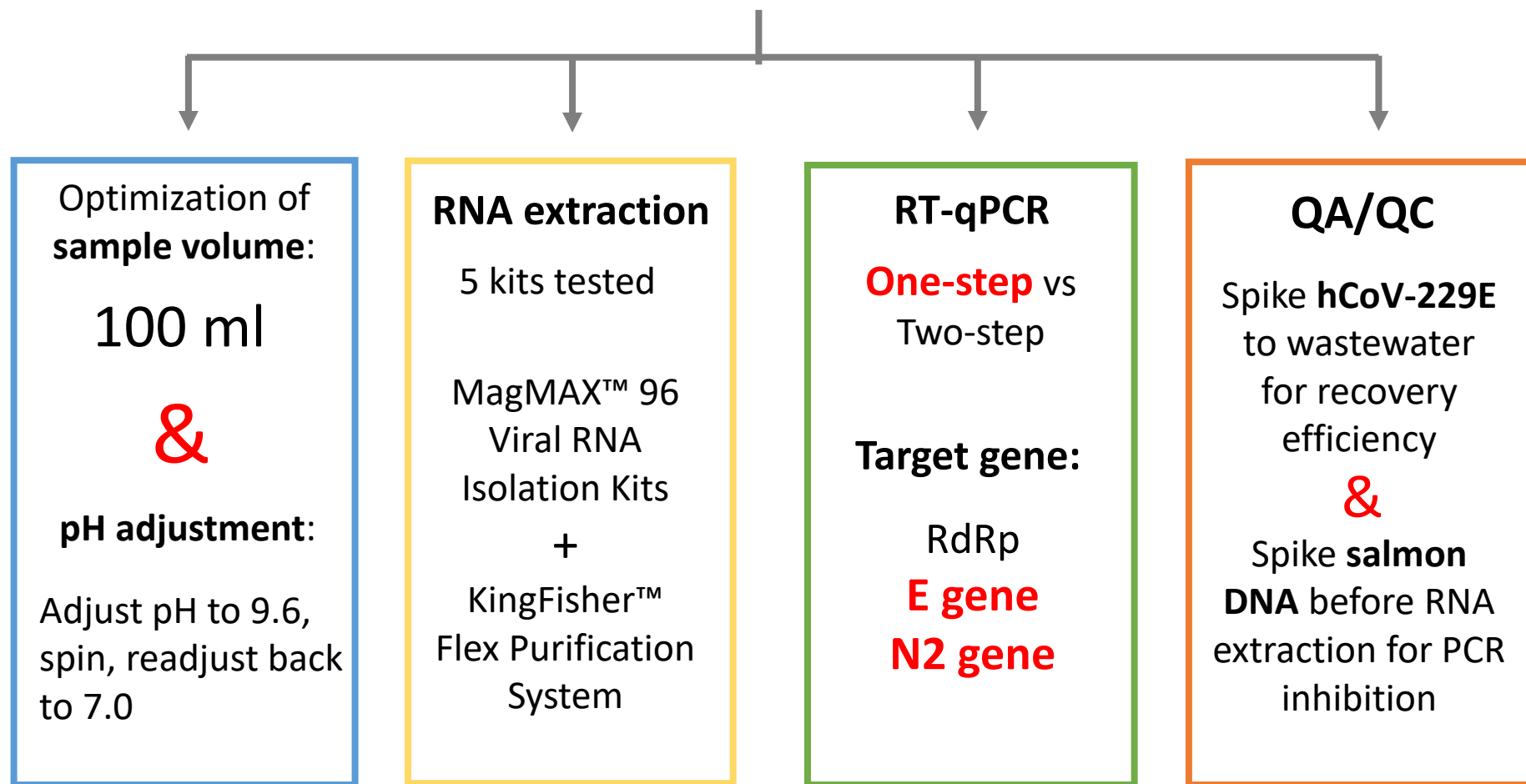
Laboratory team

Molecular Detection of SARS-CoV-2 RNA in Wastewater

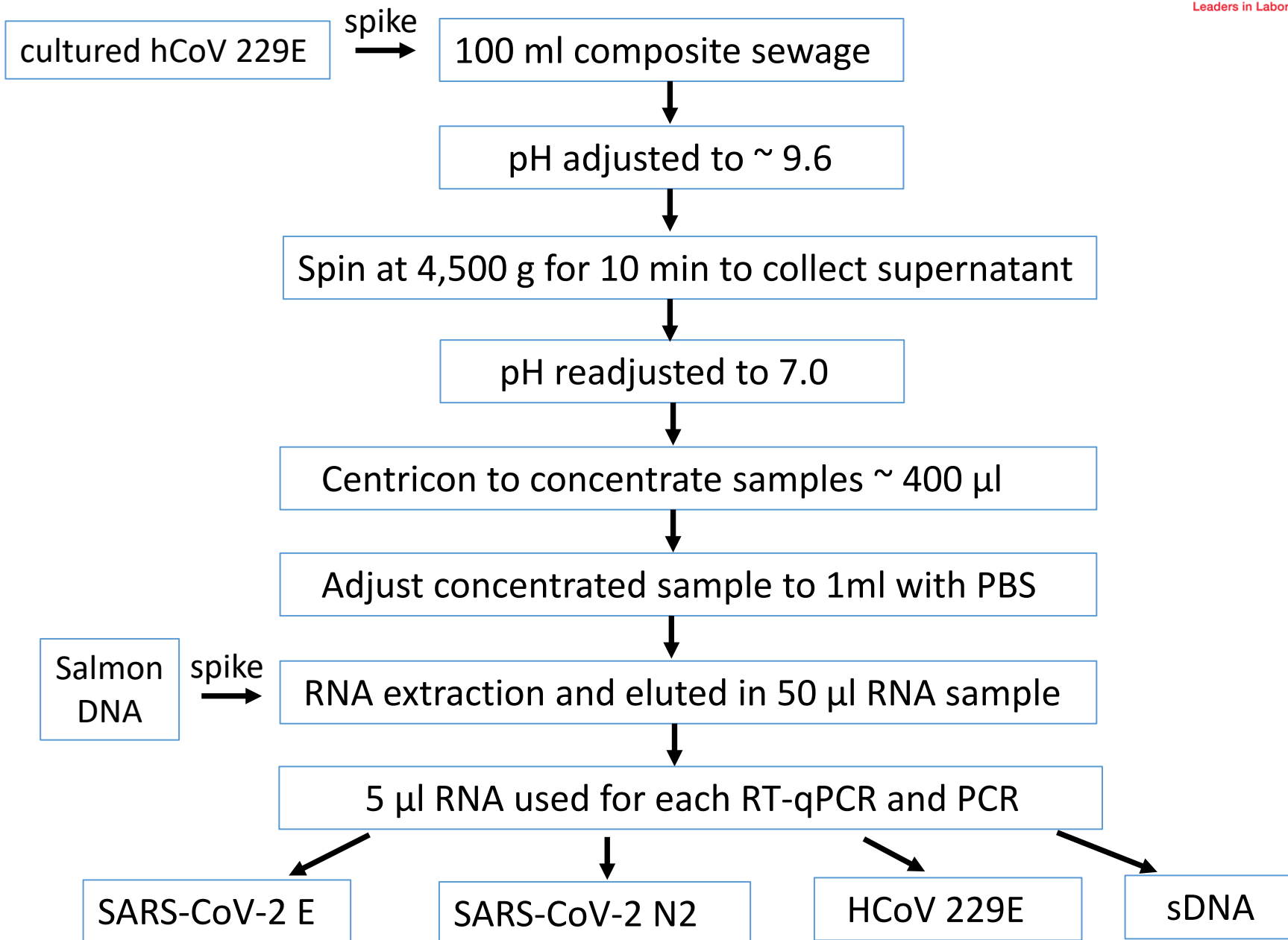
Judy Qiu & Lilly Pang

Department of Laboratory Medicine and Pathology,
University of Alberta; Public Health Laboratory
(ProvLab), Alberta Precision Laboratory

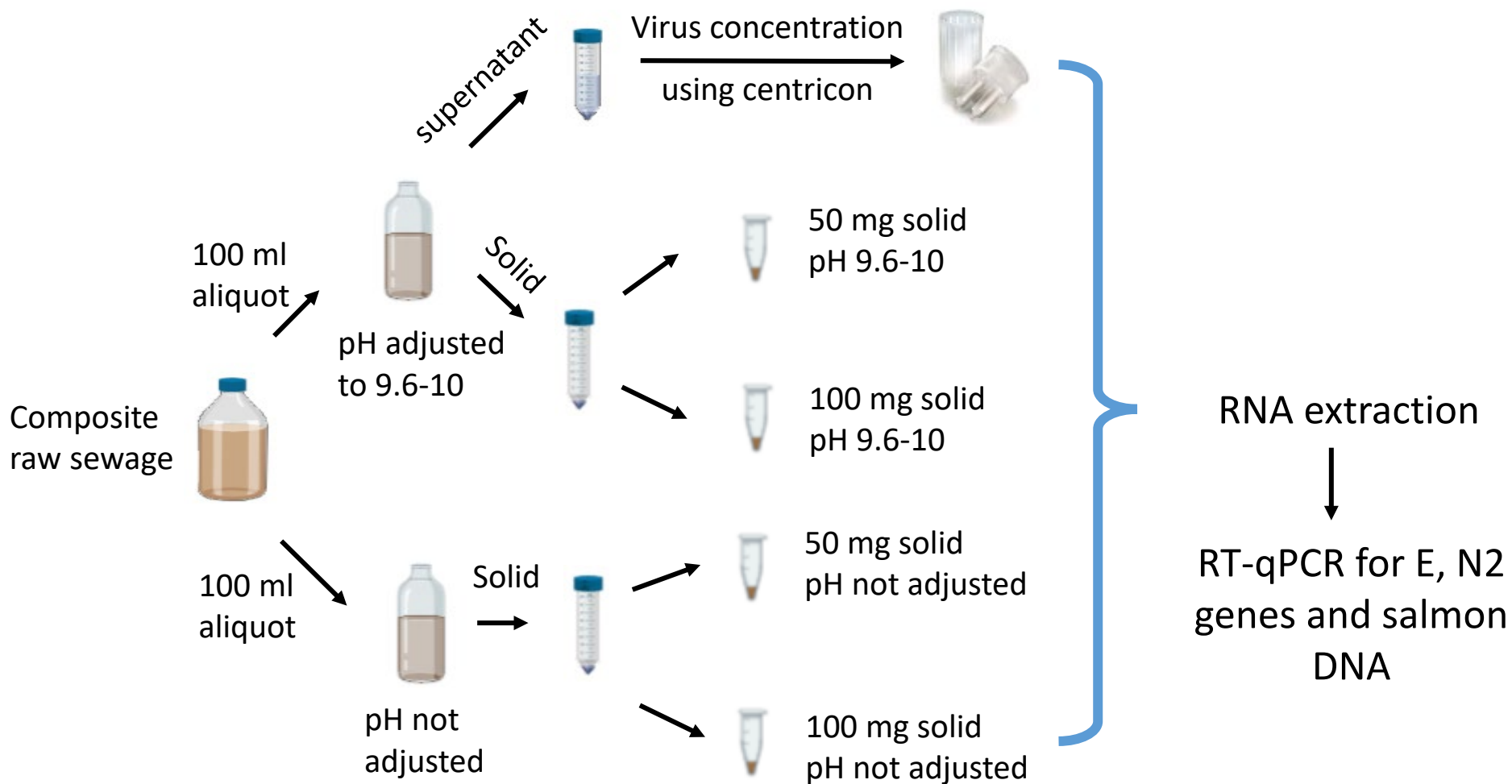
Development and Validation of Methods for Molecular Detection of SARS-CoV-2 RNA in Wastewater



Standardized laboratory protocol



Validation of Methods for Detection of SARS-CoV-2 RNA in Wastewater Solid



3		Water		Total
		+	-	
100 mg not adjusted	+	15	14	<u>29</u>
	-	11	28	39
Total		26	42	68*

*5 samples were excluded in the analysis because not enough sludge was available to test it at 100mg

5		Water		Total
		+	-	
solid	+	25	20	45
	-	1	27	28
Total		26	47	73

PCR inhibition test based on salmon DNA Ct value

Matrix type	Ct min	Ct max	Negative or Delayed (Ct over 40)	
Water (Supernatant)	26.28	37.49	10/73	(14%)
Solid 50mg/pH not adjusted	26.59	39.49	32/73	(44%)
Solid 50mg/ pH 9.6-10.0	25.97	39.91	28/72	(39%)
Solid 100mg/ pH not adjusted	26.51	39.90	38/68	(56%)
Solid 100mg/ pH 9.6-10.0	26.37	31.88	26/70	(37%)

Summary for the method of detecting SARS-CoV-2 from wastewater solids

- **Better sensitivity:** detect more SARS-CoV-2 positive samples compared to wastewater samples
- **Cost-effective:** Save the cost on centricon filter and no worries on supply shortage
- **Challenges:** Inhibition
 - Dilution
 - Increase RNA elution volume from 50 μ l to 100 μ l
- Normalized with fecal indicator PMMoV

Acknowledgements

Research team

Lilly Pang
Nicolas Ashbolt
Bonita Lee
Deena Hinshaw
Graham Tipples
James Talbot
Kimberly Simmonds
Lyndon Gyurek
Mathew Diggle
Norma Ruecker
Norman Neumann
Qiaozhi Li
Stephen Craik
Tiejun Gao

Advisory Committee

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Christopher Sikora
Debra Mooney
Graham Tipples
James Talbot
Jason Cabaj
Kathryn Koliaska
Kimberly Simmonds
Larry Svenson
Lyndon Gyurek
Norma Ruecker
Stephen Craik
S. Hrudey

Laboratory team

Lilly Pang
Judy Qiu
Eloisa Hasing
Jiao Yu
Emma Zwaigenbaum

Participating WWTP

EPCOR (Edmonton and Canmore)
ACRWC (Fort Saskatchewan)
City of Calgary
City of Red Deer
Town of High River
City of Lethbridge
City of Medicine Hat
Town of Banff
Aquatera® (Grande Prairie)



**ALBERTA PRECISION
LABORATORIES**

Leaders in Laboratory Medicine



CIHR IRSC

 Canadian Institutes of Health Research
Instituts de recherche en santé du Canada



**Alberta Health
Services**



**UNIVERSITY OF
ALBERTA**

**ALBERTA
INNOVATES** 

BCCDC's methods for detection of SARS-CoV-2 RNA in influent wastewater

Sample Preparation (4°C)

Viral Concentration (4°C)

Nucleic Acid Extraction



24-hr composite



Large particle removal



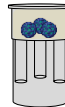
Solids (-20°C)



Liquid (filter)

**Centrifugal ultrafiltration
(30kDa MWKO)**

100mL
wastewater



~300uL
viral concentrate



**Add 1x PBS
V = 1mL**



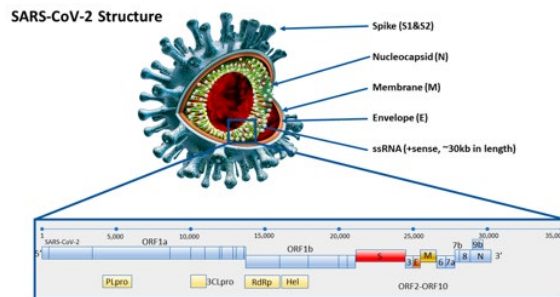
A. Automated system
NUCLISENS easyMAG
(Biomerieux)

B. Manual kit
AllPrep Power Viral
RNA/DNA kit

BCCDC's methods for detection of SARS-CoV-2 RNA in influent wastewater

SARS-CoV-2 RNA Detection

Real-Time one-step RT-qPCR



RdRP and E gene
(BCCDC PHL COVID-19 Panel)

PMMoV



Standard curves = viral copies/L

Quality Assurance and Control (QA/QC)

Limit of detection (LOD)

- 10 copies/rxn (2 copies/uL)

PCR inhibition test:

- Spike West Nile virus Armored RNA pre RNA extraction

Molecular process controls

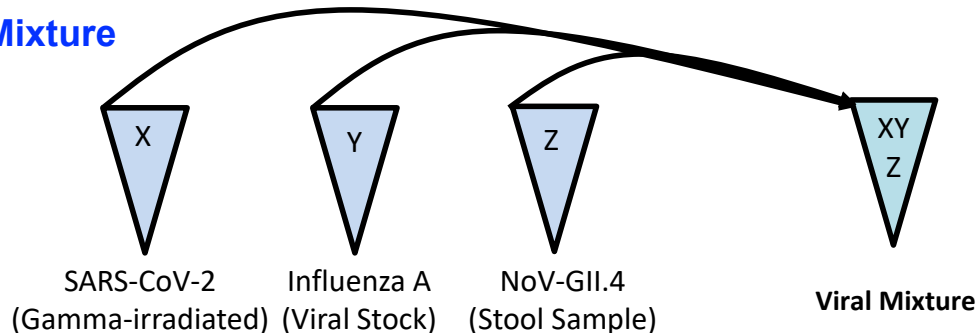
- BCCDC PHL Norovirus Panel

Viral recovery efficiency

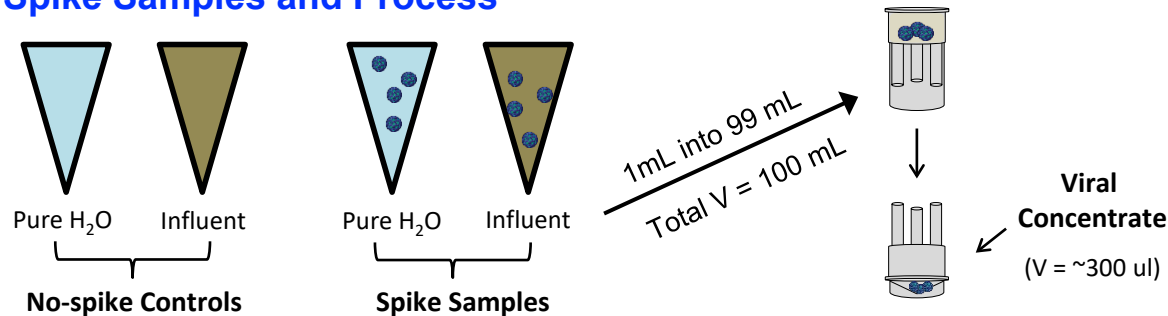
- Human influenza A
- Gamma-irradiated SARS-CoV-2
- Human norovirus GII.4
- Pending - mouse hepatitis virus

BCCDC's viral recovery workflow

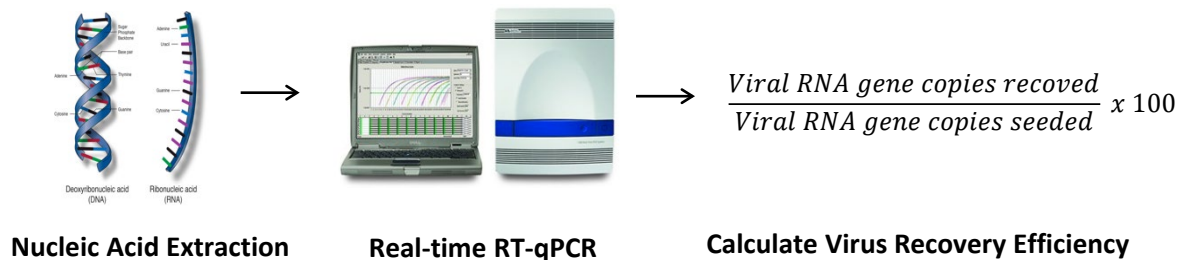
Viral Mixture



Spike Samples and Process



Viral Quantification and Recovery Efficiency



Quantify stocks by Real-time qPCR

Prepare viral mixture

Spike viral mixture into water matrices

Viral concentration

Nucleic Acid Extraction

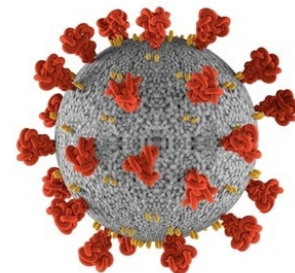
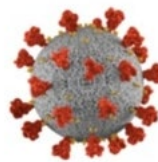
Real-time RT-qPCR w/ standard curves

Quantification

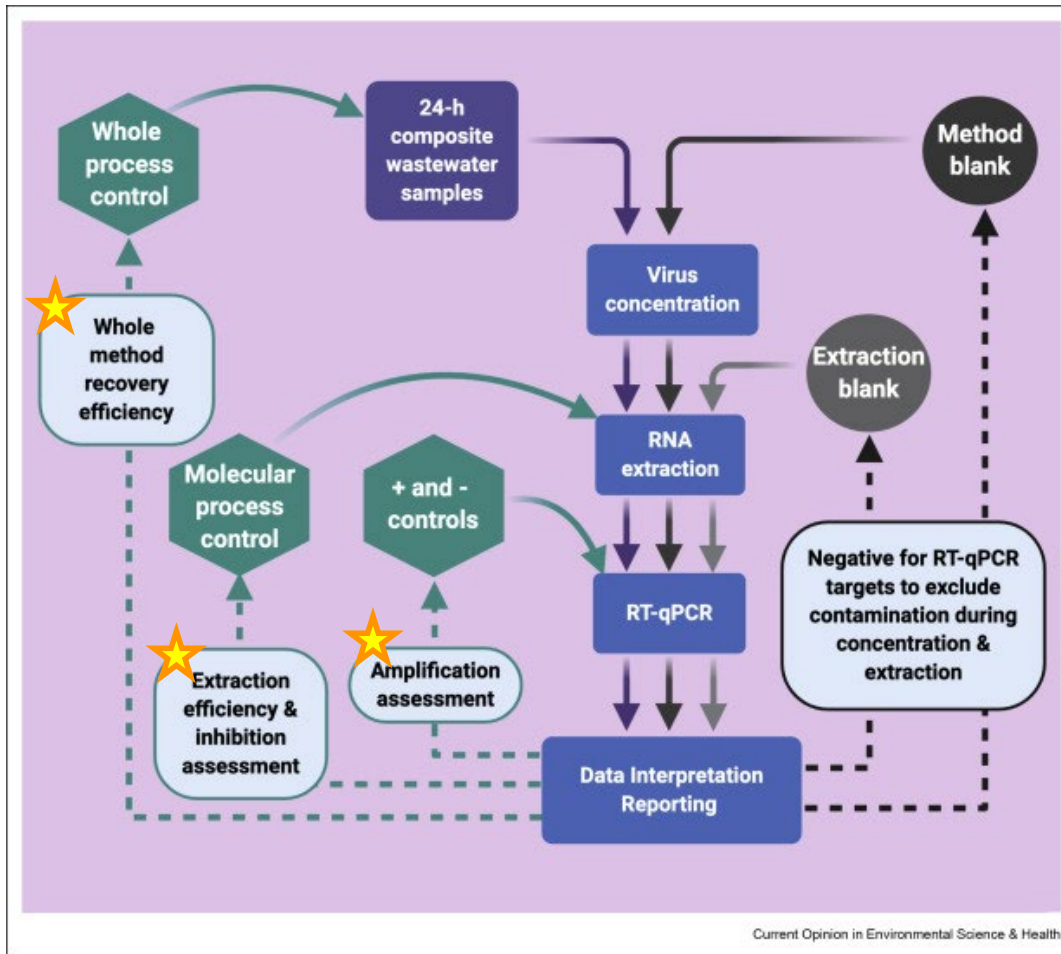
% Recovery

Measuring viral recovery is tricky

- Culturing SARS-CoV-2 requires BSL-3 laboratories and specially trained personnel
 - **SARS-CoV-2:** heat-inactivated and gamma-irradiated
- Surrogate CoV (human and nonhuman) infectious strains, or other enveloped viruses
 - **Human CoV:** 229E, OC43, SARS-CoV-1, MERS
 - **Nonhuman CoV:** MHV-A59, BCoV
 - **Human enveloped viruses:** Influenza A (H1N1)
- Must quantify your viral stock using the same method developed to quantify SARS-CoV-2 in wastewater
 - **Culture** = PFU/mL or TCID₅₀/mL
 - **RT-qPCR** = genomic copies/mL



Method optimization and QC are crucial



[Ahmed, Warish](#), et al. (2020) *Current opinion in environmental science & health*.



- Samples taken regularly, stored at 4°C and processed in 2-3 days
- Process controls should be used to evaluated viral recovery
 - whole process
 - molecular process
- Efforts should be taken to reduce the amount of inhibitors during RNA extraction
- Controls should be used to assess false negative and positive results

Acknowledgments

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- Xuan Lin



 **slack** 2019-nCoV WBE



Public Health
Agency of Canada



BC Centre for Disease Control

COVID-19 Wastewater Coalition Inter-Laboratory Study

Thank you, webinar speakers!

- Steve E. Hrudey, COVID-19 Wastewater Coalition
- Alex H.S. Chik, Inter-Laboratory Study Coordinator
- Chand Mangat, Public Health Agency of Canada – National Microbiology Laboratory
- Robert Delatolla, University of Ottawa
- Mark Servos, University of Waterloo
- Melissa Glier, BC Centre for Disease Control
- Eyerusalem Goitom, University of Ottawa
- Judy Qiu, University of Alberta



COVID-19 WASTEWATER COALITION

A national collaboration of municipal utilities, researchers, public health agencies and government with a shared goal of protecting public health from COVID-19

[COALITION UPDATES](#)[NEWS STORIES](#)[INTER-LAB STUDY](#)[REGIONAL HUBS](#)[RESOURCES](#)[GET INVOLVED](#)

Thank you for attending today's webinar series. We will reconvene at 2:00 p.m. EST for "WBE in Canada: Use cases, challenges and next steps."

Slides and recordings from the webinars will be available next week at:

cwn-rce.ca/events/webinars/cwn-webinars

Canadian COVID-19 Wastewater Coalition

Webinar series – Tuesday, December 1, 2020



Inter-Laboratory Study Outcomes & Implications

11:30 a.m. to 12:55 p.m. EST

WBE in Canada: Use cases, challenges & next steps

2:00 p.m. to 3:30 p.m. EST



CWN Webinars

Connecting water professionals to decision-ready knowledge